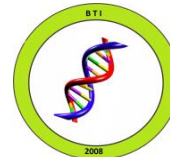




©Biotechnology Society



www.bti.org.in
ISSN 0974-1453
Review article

AN EMERGING HOPE TO COMBAT *CANDIDA ALBICANS*: PLANT BASED THERAPEUTICS

PAYAL GUPTA, PANKAJ GAUTAM, NISHANT RAI, NAVIN KUMAR*

Department of Biotechnology, Graphic Era University, Dehradun-248001, UK, India

*Corresponding Author: navinbajpai@gmail.com

ABSTRACT: *Candida* associated morbidity and mortality is reported to increase at an alarming rate worldwide due to its lead role in hospital acquired infections (Nosocomial infections). Rising incidences on resistance in *Candida* species to existing antifungal drugs and the frequent occurrence of Candidosis have prompted the researchers to look for alternative therapeutics. Several plant extracts have been screened by various groups for anti-Candidal properties. These plants could provide cost effective and safer approach of treating Candidosis, justifying the basis for developing herbal drugs against the deadly pathogen. In this review we have compiled more than hundred studies demonstrating the ability of plant extracts to combat *Candida* pathogenesis, with a perspective of developing herbal anti-*Candida* drugs.

Keywords: Anti-*Candida* Plants, Plant Extracts, Candidosis, Herbal drugs, *Candida* pathogenesis, Alternative drugs.

INTRODUCTION

Classification of *Candida*:

Kingdom: Fungi
 Phylum: Ascomycota
 Subphylum: Saccharomycotina
 Class: Saccharomycetes
 Order: Saccharomycetales
 Family: Saccharomycetaceae
 Genus: *Candida*

Candida is ubiquitous, dimorphic, opportunistic fungus that affects humans. It remains in commensal association with many animals. The infection of candida primarily depends upon the immune status of the host. *Candida* is usually diploid and naturally heterozygous yeast. *Candida albicans* displays three modes of growth: yeast, hyphae and pseudohyphae (Fig. 1), which are interconvertible depending upon the environmental conditions. These all three differ in their morphology and pathogenicity. Of all three

morphology, hyphal and pseudohyphal are pathogenic.

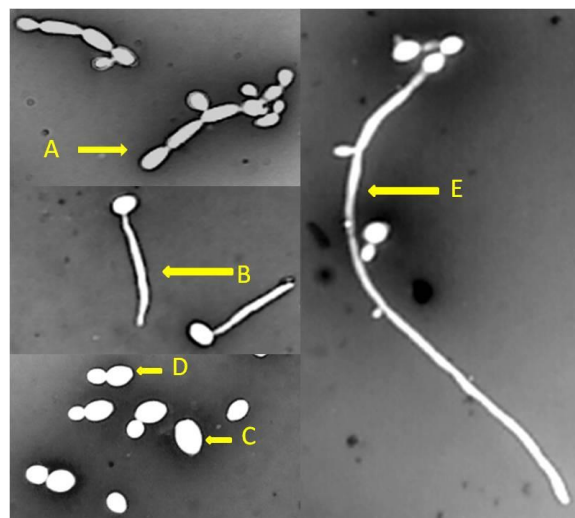


FIGURE 1: Different morphological forms of *Candida* species at 100X resolution. A- Pseudohyphae, B- Germ Tube, C- Yeast form, D- Budding form, E- Hyphae. Hyphae and pseudohyphae are induced using 10% Serum.

Now about 200 *Candida* species are known to us and of which only few are involved in human infection. More than 90% of Candidemia is caused by five *Candida* species: - *C. albicans*, *C. glabrata*, *C. tropicalis*, *C. krusei* and *C. parapsilosis* (Hajjeh *et al.*, 2004; Takakura *et al.*, 2004; Arendrup *et al.*, 2005). Candidemia may also be caused by several other non *albicans* *Candida* species like *C. dubliniensis*, *C. famata*, *C. guilliermondii*, *C. lusitaniae*, *C. norvegensis*,

C. pelliculosa and *C. rugosa* (Takakura *et al.*, 2004; Sandven *et al.*, 2008). Although these species are unusual cause of Candidiasis, but some of them are found to be associated in nosocomial clusters, or even obtained resistance to antifungal drugs (Colombo *et al.*, 2003; Colombo *et al.*, 2006).

Candida is the benign species of yeast micro-flora that initially causes a superficial mucosal infection and then with time moves deeper into bloodstreams, result into systemic infection. In its hyphal form mainly, it invades the epithelial tissue and penetrates deeper into tissues. After *C. albicans*, *C. glabrata* is the second or third most cause of Candidiasis and is not dimorphic in nature.

Based on the part of the body where infection has occurred, the main types of *Candida* infections are systemic Candidiasis, oral thrush, oesophageal thrush, cutaneous Candidiasis, *Candida* vaginitis and *Candida* balanitis. Blood stream infection of *Candida* is called Candidemia that is the most frequent clinical manifestation

and causes significant morbidity and mortality in hospitalized patients.

Candida species is the fourth most frequent cause for nosocomial systemic infection in ICU patients. Also, this yeast is the third most widespread cause of nosocomial bloodstream infections (Alimirante *et al.*, 2005). Various factors contribute towards such a high rate of this infection like lack of resource for medical care and training programs, difficulties in controlling the infection in hospitals in developing countries, shortage of staff in healthcare and less effective anti-fungal drugs. Mostly cases of *Candida* infection reported include areas of neonatal, paediatric and adult-ICUs because these persons have suppressed immune responses against *Candida*. The three major groups of patients that are at high risk of Candidiasis are with neutropenic cancer, organ or stem cell transplant and those undergoing intensive care procedures. In past few years, it has been observed that the patients having a central venous catheter, receiving parental nutrition or on broad-spectrum

antibiotics are also at high risk. They have high acute physiology and chronic health evaluation scores (Rangel-Frausto *et al.*, 1999; Michalopoulos *et al.*, 2003; Pappas *et al.*, 2003). It causes severe infection ranging from superficial muco-cutaneous to tissue invasive. A limited number of drugs are available against it. This problem is contributed due to the factors like dimorphic transition, adhesion, invasion, resistance to phagocytosis and ability to grow well at 37°C. Of many species, *Candida albicans* receive special attention because of its association with humans.

The exact mechanism involved in candidiasis is not known but it is clear that there must be some imbalance occurs between host-pathogen relationships that must be in favour of pathogen (Chakravarthi *et al.*, 2010). According to geographical settings and demographics of the population studied, the frequency of systemic Candidiasis varies in blood samples. Due to the large use of fluconazole drugs for the treatment of HIV-infected patients having candidiasis results

in the less drug resistant strains of the yeast in early 1990s. These resistant strains emerged out because of their acquisition with previously sensitive *C. albicans* (Sanglard *et al.*, 2002). But according to the information gathered from Population-based and sentinel surveillance programs worldwide, it was observed that frequency of fluconazole resistance among *C. albicans* remains insignificant (Hajjeh *et al.*, 2004; Alimirante *et al.*, 2005; Colombo *et al.*, 2006).

Another new thing was noted in numerous hospitals that the frequency of *C. glabrata* is enhanced as a serious *Candida* infection (Baddley *et al.*, 2001; Malani *et al.*, 2001). Reasons for this include living habitat, age and type of the patients and use of fluconazole. Also *C. glabrata* infection is mainly noted in persons more than 60 years (Malani *et al.*, 2001) and in those who have leukaemia or received stem cell transplant and those who are associated with increasing use of fluconazole (Marr *et al.*, 2000).

CURRENT STATUS OF CANDIDOSIS

Candida infections are increasing alarmingly in India. According to a report, the most common are *C. albicans* (47%) followed by *C. Tropicalis* (23%). In one of the study at a multi super-speciality centre in southern India, it is found that 74% of incidences of Candidaemia have been due to non-*albicans Candida*. Of which *C. tropicalis* accounted for 39.7% of cases, followed by *C. albicans* (26.4%), *C. lipolytica* (5.8%), *C. famata* (4.4%), *C. parapsilosis* (4.4%), *C. pelliculosa* (4.4%), *C. Guilliermondii* (2.9%), *C. Lusitaniae* (1.4%), *C. sphaeria* (1.4%) and other *Candida* species (8.8%). A study done in the Tertiary Care Institute in India (BL Kapur Memorial Hospital, Delhi) suggested that 18% of all nosocomial blood infection is caused by *Candida* species and reported 53 incidence of Candidaemia in 48 patients in 17 months. They found that 45% of incidences were due to *C. tropicalis*, 23% by *C. albicans* and 32% by other *Candida* species. The study of susceptibility profile of *Candida*

isolates showed 100% sensitivity to voriconazole, 92% to amphotericin B, 90% to flucytosine and 75% to fluconazole. But a study from India suggested that candida isolates has shown a very high resistance to voriconazole (56%), fluconazole (36%), itraconazole (24%), but shown susceptibility of 92% to amphotericin B and 96% to flucytosine. While according to a report from western India, *C. albicans* is associated with 50-70% of all cases of Candidaemia (Report by A. Kothari and V. Sagar, 2009 and Brief communication by R. Adhikary and S. Joshi, 2011).

In US, Candidemia infection is the fourth most widely hospital acquired infection (HAI) (Wisplinghoff *et al.*, 2004). According to a report (Hajjeh *et al.*, 2004), frequency of Candidal HAI in US is 1.5 cases per 10,000 patients days, whereas in Europe, frequency is slightly lower to 0.5-0.7 cases per 10,000 patients days (Richet *et al.*, 2002; Marchetti *et al.*, 2004). In 2006, the highest cases of Candidemia infection reported from Brazil, i.e.,

3.7 cases per 10,000 patients days (Colombo *et al.*, 2006).

In US, from 1997 to 2001 mortality rate was about 49% for Candidemia patients in medical centres (Gudlaugsson *et al.*, 2003) that was higher than earlier by 11% from 1983 to 1988 (Wey *et al.*, 1988). According to some of the analysis, incidences of Candidemia get complicated with time. The major cases of Candidemia results from *Candida albicans* that mainly causes infection in immune-competant patients. This rate, in Netherland, rose from 0.37 to 0.72 cases per 10,000 patients days between 1987 to 1995 (Voss *et al.*, 1996). While in Switzerland, the rate remains same approximately between 0.5 cases per 10,000 patient day from 1991 to 2000 (Fidel, 2006).

Various factors contribute towards such a high rate of this infection like lack of resource for medical care and training programs, difficulties in controlling the infection in hospitals in developing countries, shortage of staff in healthcare and less effective anti-fungal drugs.

POSSIBILITIES WITH PLANT BASED THERAPEUTICS

From the time immemorial, the plants are serving humans in several ways. The use of plant resources for medicinal and other purposes is one of a number of practices developed by ancient people. Most of the world's population relies on herbal medicine (Lewington *et al.*, 2003). In ancient times, plants used randomly without knowing the exact compound that are responsible for its therapeutic responses. But nowadays, as the advances going in herbal medicines, the active chemical in plants are now known to us and their efficacy is also known. In most developing countries, the indigenous modes of herbal treatment are a part of the culture and the dominant method of healing therapy. Reviews have compiled modern researches, which have proven the ability of plant extracts against different pathophysiological conditions and pathogenic infections (Nagle and Zohu, 2006; Anand *et al.*, 2012; Rai *et al.*, 2012; Joshi *et al.*,

2013; Ruwali *et al.*, 2013; Kumar *et al.*, 2013). These remedies, with a considerable extent of effectiveness, are socially accepted, economically viable and, mostly, are the only available source. WHO has listed 20,000 medicinal plants used in different parts of the world. Other estimates indicate the number to range between 35,000 and 70,000 worldwide (Lewington *et al.*, 2003; Bhattarai *et al.*, 2004). Several medicinal systems of the world are dependent upon plants like-Chinese, Tibetan, Indian (Ayurveda), Unani, Siddha and Homeopathy systems. Many countries, especially in Asia, Africa and East European countries, have officially recognized the use of plant-based traditional medicine in their health care delivery systems. WHO has estimated

that 80% of the world's populations rely primarily on traditional medicine (WHO Technical Report Series, No. 622:8, Geneva, Switzerland; Okerele, 1992). Considering the coverage and effectiveness of various systems of traditional medicine throughout the world, the Alma-Ata Declaration of the WHO in 1978 proposed the theme “**Health for all by the year 2000**”.

These plants provide a way for treating those pathogenic diseases against which no drug is available or for which pathogens are becoming resistant. In the present article, we are providing the information about more than 100 plants and their different extracts, showing anti-*Candida* properties (specifically against *C. albicans*) that have been screened till now, all over the globe.

Table 1: PLANTS SHOWING ANTI-CANDIDA PROPERTIES ALONG WITH THEIR PHYTOCONSTITUENTS AND OTHER RELATED PARAMETERS AND SOURCE REFERENCES.

Sr. No.	Plant (Family) [Reference]	Plant Part	Extraction Solvent	Phytochemical constituents	ZOI (mm)	MIC ₅₀ µg/ml	MFC ₅₀ µg/ml
1	<i>Aloe vera</i> (Xanthorrhoeaceae) [Yebpella <i>et al.</i> , 2011;	Leaf pulp,	Methanol	(Anthraquinones/ anthrones, carbohydrates, chromes, inorganic &	18-25	NI	NI

	Hamman <i>et al.</i> , 2008*; Ibrahim <i>et al.</i> , 2011]	Green grid	Ethanol Acetone	organic compound, amino acids, vitamins and saccharides.)* Flavonoids, tannins, steroid	9.96 10.20	NI NI	NI NI
2	<i>Sapindus saponaria</i> (Sapindaceae) [Damke <i>et al.</i> , 2011; Tzuzuki <i>et al.</i> , 2007*]	Fruit pericarp	Hydroalcohol butanol	(Triterpene saponin- S1, S2 & oligoglycoside- 1(OGSA-1))*)	NI NI	9 36	310 150
3	<i>Punica granatum</i> (Lythraceae) [Endo <i>et al.</i> , 2010; Hoffling <i>et al.</i> , 2010; Pai <i>et al.</i> , 2010; Beckman, 2007*]	Fruit peel Fruit Fruit	Hydro- alcoholic Dichloro- methane Methanol Aqueous	Punicalagin NI NI (Tannins)*	NI NI NI 22-18.8	3.9 1 3 NI	NI NI NI NI
4	<i>Acacia nilotica</i> (Fabaceae) [Khan <i>et al.</i> , 2009; Pai <i>et al.</i> , 2010]	Bark Leaves	Aqueous Ethanol	Tannins NI	4.62-7.62 26-35	NI 4.9	NI 19.5
5	<i>Cuminum cyminum</i> (Apiaceae) [Khan <i>et al.</i> , 2009]	Dried seeds	Diethyl ether	Oil	1.81-6.5	NI	NI
6	<i>Cinnamomum verum</i> (Lauraceae) [Ababutain <i>et al.</i> , 2011; Bakkali <i>et al.</i> , 2005*; Redding <i>et al.</i> , 1993*; Saad <i>et al.</i> , 2005*]	Bark	ethanol	[Phenol (eugenol), tannins, flavonoids]*	3.8	25	-
7	<i>Syzygium aromaticum</i> (Myrtaceae) [Ababutain <i>et al.</i> , 2011; Bakkali <i>et al.</i> , 2005*; Redding <i>et al.</i> , 1993*; Saad <i>et al.</i> , 2005*; Khan <i>et al.</i> , 2009; Hammer <i>et al.</i> , 1999]	Buds Buds Bud	Ethanol Ethanol AC	Phenol (Eugenol)* Aldehyde (Cinnamaldehyde) Essential oil	4.8 >40 NI	12.5 156 0.12	NI 156 NI
8	<i>Vitex negundo</i> (Verbenaceae) [Kumar <i>et al.</i> , 2011]	Leaf	Aqueous Ethanol Methanol	NI NI NI	6-5 3 2.5-0.5	NI NI NI	NI NI NI
9	<i>Adathoda vasica</i> (Acanthaceae) [Kumar <i>et al.</i> , 2011]	Leaf	Aqueous Ethanol Methanol	NI NI NI	7-4 5-3 2-1	NI NI NI	NI NI NI

10	<i>Azadirachta indica</i> (Meliaceae) [Kumar <i>et al.</i> , 2011; Panda <i>et al.</i> , 2012]	Leaf	Aqueous	NI	3.5-2	NI	NI
			Ethanol	NI	2	NI	NI
			Methanol	NI	3-1	NI	NI
		Leaves	Benzene				
			Chloroform	NI	13	26.3	NI
			Ethanol	NI	16	13.15	NI
				NI	10	52.6	NI
11	<i>Mentha piperita</i> (Lamiaceae) [Kumar <i>et al.</i> , 2011; Hammer <i>et al.</i> , 1999; Sartoratto <i>et al.</i> , 2004]	Leaf	Aqueous	NI	4-3	NI	NI
			Ethanol	NI	2-1	NI	NI
			Methanol	NI	2-1.5	NI	NI
		Herb	AC	Essential oil	NI	0.5	NI
		Aerial part	AC	Essential oil 3-octanol, isopentyl- n-butyrate, linalool, <i>cis-p</i> -menth-2-en-1- ol, terpin-4-ol, α - terpineol, carvone, β - bourbonene, <i>trans</i> - caryophyllene, germacrene D,	NI	0.74	NI
12	<i>Curcuma longa</i> (Zingiberaceae) [Kumar <i>et al.</i> , 2011]	rhizome	Aqueous	NI	2.5	NI	NI
			Ethanol	NI	3.5-1	NI	NI
			Methanol	NI	5-3	NI	NI
13	<i>Matricaria chamomillathyme</i> (Asteraceae) [Dalirsani <i>et al.</i> , 2011]	Herb	Methanolic	NI	18.98	NI	NI
14	<i>Allium vineale</i> (Amaryllidaceae) [Dalirsani <i>et al.</i> , 2011; Xia <i>et al.</i> , 2005*; Ankri <i>et al.</i> , 1999*; Wang <i>et al.</i> , 2000*; Wang <i>et al.</i> , 2001*]		Methanolic	(Alliumin, allicin, allivin, lillin)*	18.64	NI	NI
15	<i>Cinnamomum zeylanicum</i> (Lauraceae) [Dalirsani <i>et al.</i> , 2011; Singh <i>et al.</i> , 1995*; Khan <i>et al.</i> , 2009]	Bark	Methanolic	(Cinnamic aldehyde)*	14.92	NI	NI
			Ethanol	NI	>40	19.5	78
16	<i>Eugenia aromatic</i> (Myrtaceae) [Dalirsani <i>et al.</i> , 2011]		Methanolic	NI	15.26	NI	NI

17	<i>Salvia officinalis</i> (Lamiaceae) [Dalirsani <i>et al.</i> , 2011; Hammer <i>et al.</i> , 1999]	Herb	Methanolic AC	NI Essential oil	14.8 NI	NI 0.5	NI NI
18	<i>Thymus serpyllum</i> (Lamiaceae) [Dalirsani <i>et al.</i> , 2011]		Methanolic	NI	8.81	NI	NI
19	<i>Momordica charantia</i> (Cucurbitaceae) [Mwambete <i>et al.</i> , 2009]	Fruit	Methanol Petroleum ether	Momordin, α - & β - momocharin, cucurbitacin B1 and oleanolic acid	10 8	NI	NI
20	<i>Cassia occidentalis</i> (Fabaceae) [Arya <i>et al.</i> , 2010]	Leaf	Methanol aqueous	NI NI	8 5	NI NI	NI NI
21	<i>Berchemia discolor</i> (Rhamnaceae) [Samei <i>et al.</i> , 2010]	Leaves Bark	Hexane Acetone Hexane Acetone	NI	12-10 - - -	3.75 0.23 7.5 3.75	7.5 - 7.5 -
22	<i>Cassine transvaalensis</i> (Celastraceae) [Samei <i>et al.</i> , 2010]	Bark	Hexane	NI	16-12	0.46	3.75
23	<i>Diospyros mespiliformis</i> (Ebenaceae) [Samei <i>et al.</i> , 2010]	Leaves	Hexane	NI	7	-	-
24	<i>Piper capense</i> (Piperaceae) [Samei <i>et al.</i> , 2010]	Root	Acetone Hexane	NI	24-22 12-8	1.88 3.75	3.75 7.5
25	<i>Warburgia salutaris</i> (Canellaceae) [Samei <i>et al.</i> , 2010]	Bark Leaves	Acetone Hexane Acetone Hexane	NI	22-7 16-10 15 22-16	3.75 0.23 7.5 7.5	7.5 0.93 0.0 -
26	<i>Anethum graveolens</i> (Apiaceae) [Zeng <i>et al.</i> , 2011]	Leaves	Aqueous (Steam)	Essential oil (Carvone and limonene)	NI	0.625	NI
27	<i>Foeniculum vulgare</i> (Apiaceae) [Pai <i>et al.</i> , 2010]	Seeds	Diethyl-ether	NI	2.5	NI	NI
28	<i>Pisidium guajava</i> (Myrtaceae) [Jebashree <i>et al.</i> , 2011; Suvarna <i>et al.</i> , 2009]	Leaves	Hexane Ethyl acetate Ethanol Methanol Aqueous Methanol	NI NI	16 16 15 15 11.3 11.7	NI NI NI	NI NI NI

			Chloroform Petroleum- ether		12.7 10.3	NI	NI
29	<i>Terminalia chebula</i> (Combretaceae) [Jebashree <i>et al.</i> , 2011]	Dried fruits	Hexane Ethyl acetate Methanol	NI	12 16 11	NI NI NI	NI NI NI
30	<i>Achyranthes aspera</i> (Amaranthaceae) [Jebashree <i>et al.</i> , 2011]	Stem	Ethyl acetate Ethanol Methanol	NI	15 12 13	NI NI NI	NI NI NI
31	<i>Rosmarinus officinalis</i> (Lamiaceae) [Hoffling <i>et al.</i> , 2010; Hammer <i>et al.</i> , 1999]	Leaves	Dichloro- methane Methanol Essential oil	NI NI	NI NI	7 1	NI NI
		Herb		NI	NI	1	NI
32	<i>Arrabidaea chica</i> (Bignoniaceae) [Hoffling <i>et al.</i> , 2010]	Leaves	Dichloro- methane Methanol	NI NI	NI NI	15 7	NI NI
33	<i>Tabebuia avellanadae</i> (Bignoniaceae) [Hoffling <i>et al.</i> , 2010]	Bark	Dichloro- methane Methanol	NI NI	NI NI	Resistant 3	NI NI
34	<i>Syzygium cumini</i> (Myrtaceae) [Hoffling <i>et al.</i> , 2010]	Seeds	Dichloro- methane Methanol	NI NI	NI NI	30 1	NI NI
35	<i>Cassia tora</i> (Caesalpinaceae) [Panda <i>et al.</i> , 2012; Maji <i>et al.</i> , 2010]	Leaves	Benzene Chloroform Ethanol Water Acetone Benzene	NI NI NI NI NI NI	10 10 16 4 6 5	26.3 13.15 26.3 - - -	NI NI NI - - -
36	<i>Lawsonia inermis</i> (Lythraceae) [Panda <i>et al.</i> , 2012]	Leaves	Benzene Chloroform Ethanol	NI NI NI	7 7 11	26.3 13.15 52.6	NI NI NI
37	<i>Pseudolachnostylis maprouneifolia</i> (Phyllanthaceae) [Panda <i>et al.</i> , 2012]	Leaves	Hexane Acetone	NI	10-0 0.0	>7.5 1.88	7.5 0.0
38	<i>Lipia javanica</i> (Verbenaceae) [Panda <i>et al.</i> , 2012]	Leaves	Hexane Acetone	NI	10-0 -	3.75 >7.5	- -

39	<i>Maeruae dulis</i> (Capparaceae) [Panda <i>et al.</i> , 2012]	Root	Hexane Acetone	NI	- -	3.75 1.88	- -
40	<i>Bauhinia galpinii</i> (Fabaceae) [Panda <i>et al.</i> , 2012]	Bark Leaves	Hexane Hexane Acetone	NI NI NI	- - -	1.88 1.88 0.23	- - -
41	<i>Bridelia micrantha</i> (Phyllanthaceae) [Panda <i>et al.</i> , 2012]	Bark	Acetone Hexane	NI NI	- -	>7.5 3.75	- -
42	<i>Bridelia mollis</i> (Euphorbiaceae) [Panda <i>et al.</i> , 2012]	Leaves	Acetone Hexane	NI NI	- -	3.75 >7.5	- -
43	<i>Cassia petersiana</i> (Fabaceae) [Panda <i>et al.</i> , 2012]	Bark	Acetone	NI	-	>7.5	-
44	<i>Capparis tomentosa</i> (Capparaceae) [Panda <i>et al.</i> , 2012]	Root	Acetone	NI	-	3.75	-
45	<i>Pouzolzia mixtra</i> (Urticaceae) [Panda <i>et al.</i> , 2012]	Leaves	Acetone Hexane	NI	-	>7.5 3.75	-
46	<i>Rhoicissus tridentate</i> (Vitaceae) [Panda <i>et al.</i> , 2012]	Tubes	Acetone Hexane	NI	-	>7.5 3.75	-
47	<i>Schotia barchypet ala</i> (Fabaceae) [Panda <i>et al.</i> , 2012]	Bark	Hexane	NI	-	3.75	-
48	<i>Strychnos decussate</i> (Loganiaceae) [Panda <i>et al.</i> , 2012]	Bark	Acetone Hexane	NI	-	3.75 3.75	-
49	<i>Senna didymobotrya</i> (Fabaceae) [Panda <i>et al.</i> , 2012]	Root	Acetone Hexane	NI	-	7.5 7.5	-
50	<i>Terminalia sericea</i> (Combretaceae) [Panda <i>et al.</i> , 2012]	Root	Acetone	NI	-	1.88	-
51	<i>Mangifera indica</i> (Anacardiaceae) [Suvarna <i>et al.</i> , 2009]	Leaves	Aqueous Methanol Chloroform Petroleum- ether	NI NI NI NI	7.7 10.7 6.3 5.7	NI NI NI NI	NI NI NI NI
52	<i>Ixora parviflora</i> (Rubiaceae) [Suvarna <i>et al.</i> , 2009]	Leaves	Aqueous Methanol Chloroform Petroleum-	NI NI NI NI	11.3 10.7 9	NI NI NI	NI NI NI

			ether	NI	13	NI	NI
53	<i>Calotropis procera</i> (Asclepiadaceae) [Suvarna <i>et al.</i> , 2009]	Leaves	Chloroform	NI	6.3	NI	NI
54	<i>Syzygium jambolanum</i> (Myrtaceae) [Prabhakar <i>et al.</i> , 2008]	Bark	Ethanol	NI	19-22	NI	NI
55	<i>Cassia siamea</i> (Fabaceae) [Prabhakar <i>et al.</i> , 2008]	Bark	Ethanol	NI	19-22	NI	NI
56	<i>Odina wodier</i> (Anacardiaceae) [Prabhakar <i>et al.</i> , 2008]	Bark	Ethanol	NI	11-14	NI	NI
57	<i>Sargassum wightii</i> (Sargassaceae) [Prabhakar <i>et al.</i> , 2008]	Complete body	Ethanol	NI	19-22	NI	NI
58	<i>Caulerpa scalpelliformis</i> (Caulerpaceae) [Prabhakar <i>et al.</i> , 2008]	Complete body	Ethanol	NI	19-22	NI	NI
59	<i>Aniba rosaedora</i> (Lauraceae) [Hammer <i>et al.</i> , 1999]	Wood	AC	Essential Oil	NI	0.25	NI
60	<i>Apium Graveolens</i> (Apiaceae) [Hammer <i>et al.</i> , 1999]	Seed	AC	Essential Oil	NI	1	NI
61	<i>Boswellia carterii</i> (Burseraceae) [Hammer <i>et al.</i> , 1999]	Resin	AC	Essential Oil	NI	1	NI
62	<i>Cananga odorata</i> (Annonaceae) [Hammer <i>et al.</i> , 1999]	Flower	AC	Essential Oil	NI	1	NI
63	<i>Cedrus atlantica</i> (Pinaceae) [Hammer <i>et al.</i> , 1999]	Wood	AC	Essential Oil	NI	>2	NI
64	<i>Citrus aurantifolia</i> (Rutaceae) [Hammer <i>et al.</i> , 1999]	Fruit	AC	Essential Oil	NI	2	NI
65	<i>Citrus aurantium</i> (Rutaceae) [Hammer <i>et al.</i> , 1999]	Peel	AC	Essential Oil	NI	1	NI
66	<i>Citrus lemon</i> (Rutaceae) [Hammer <i>et al.</i> , 1999]	Peel	AC	Essential Oil	NI	2	NI
67	<i>Citrus x paradise</i> (Rutaceae)	Peel	AC	Essential Oil	NI	1	NI

	[Hammer <i>et al.</i> , 1999]						
68	<i>Citrus reticulata</i> var. <i>Madurensis</i> (Rutaceae) [Hammer <i>et al.</i> , 1999]	Peel	AC	Essential Oil	NI	2	NI
69	<i>Commiphora myrrha</i> (Burseraceae) [Hammer <i>et al.</i> , 1999]	Resin	AC	Essential Oil	NI	>2	NI
70	<i>Coriandrum sativum</i> (Apiaceae) [Hammer <i>et al.</i> , 1999]	Seed	AC	Essential Oil	NI	0.25	NI
71	<i>Cucurbita pepo</i> (Cucurbitaceae) [Hammer <i>et al.</i> , 1999]	Seed	AC	Essential Oil	NI	>2	NI
72	<i>Cupressus sempervirens</i> (Cupressaceae) [Hammer <i>et al.</i> , 1999]	Leaves & twigs	AC	Essential Oil	NI	>2	NI
73	<i>Cymbopogon citratus</i> (Poaceae) [Hammer <i>et al.</i> , 1999]	Leaf	AC	Essential Oil	NI	0.06	NI
74	<i>Cymbopogon martini</i> (Poaceae) [Hammer <i>et al.</i> , 1999]	Leaf	AC	Essential Oil	NI	0.06	NI
75	<i>Cymbopogon nardus</i> (Poaceae) [Hammer <i>et al.</i> , 1999]	Leaf	AC	Essential Oil	NI	0.12	NI
76	<i>Daucus carota</i> (Apiaceae) [Hammer <i>et al.</i> , 1999]	Seed	AC	Essential Oil	NI	2	NI
77	<i>Eucalyptus polybractea</i> (Myrtaceae) [Hammer <i>et al.</i> , 1999]	Leaves & twigs	AC	Essential Oil	NI	1	NI
78	<i>Foeniculum vulgare</i> (Apiaceae) [Hammer <i>et al.</i> , 1999]	Seed	AC	Essential Oil	NI	0.5	NI
79	<i>Gaultheria procumbens</i> (Ericaceae) [Hammer <i>et al.</i> , 1999]	Herb	AC	Essential Oil	NI	0.25	NI
80	<i>Juniperus communis</i> (Cupressaceae) [Hammer <i>et al.</i> , 1999]	Berry	AC	Essential Oil	NI	2	NI
81	<i>Lavandula angustifolia</i> (Lamiaceae) [Hammer <i>et al.</i> , 1999]	Flower	AC	Essential Oil	NI	0.5	NI
82	<i>Macadamia integrifolia</i> (Proteaceae) [Hammer <i>et al.</i> , 1999]	Nut	AC	Essential Oil	NI	>2	NI

83	<i>Melaleuca alternifolia</i> (Myrtaceae) [Hammer <i>et al.</i> , 1999]	Leaves & twigs	AC	Essential Oil	NI	0.5	NI
84	<i>Melaleuca cajuputi</i> (Myrtaceae) [Hammer <i>et al.</i> , 1999]	Leaves & twigs	AC	Essential Oil	NI	1	NI
85	<i>Melaleuca quinquenervia</i> (Myrtaceae) [Hammer <i>et al.</i> , 1999]	Leaves & twigs	AC	Essential Oil	NI	0.25	NI
86	<i>Mentha spicata</i> (Lamiaceae) [Hammer <i>et al.</i> , 1999; Sartoratto <i>et al.</i> , 2004]	Herb	AC	Essential Oil: Piperitenone oxide, β - farnesene, γ - muurolene,	NI NI	0.12 >2.00	NI NI
87	<i>Ocimum basilicum</i> (Lamiaceae) [Hammer <i>et al.</i> , 1999; Sartoratto <i>et al.</i> , 2004]	Herb	AC	Essential Oil: 1,8-cineol, fenchone, linalool, camphor, terpin-4-ol, α - terpineol, eugenol, β - bourbonene, β - elemene, <i>trans</i> - caryophyllene, β - bergamotene, α - guaiene, α -humulene, germacrene D, δ - guaiene, δ -cardiene, γ -cardiene, epi- α - muurolol, α - eudesmol, α -cadinol.	NI NI	0.5 >2.00	NI NI
88	<i>Ocimum gratissimum</i> (Lamiaceae) [Sartoratto <i>et al.</i> , 2004]	Herb	AC	Essential Oil : Terpin-4-ol, <i>trans</i> - caryophyllene, eugenol, germacrene D, δ -cardiene, α - cadinol.	NI	>2.0	NI
89	<i>Oenothera biennis</i> (Onagraceae) [Hammer <i>et al.</i> , 1999]	Seed	AC	Essential Oil	NI	>2	NI
90	<i>Origanum majorana</i> (Lamiaceae) [Hammer <i>et al.</i> , 1999]	Herb	AC	Essential Oil	NI	0.25	NI
91	<i>Origanum vulgare</i> (Lamiaceae) [Hammer <i>et al.</i> , 1999; Sartoratto <i>et al.</i> , 2004]	Herb	AC	Essential Oil: 1-octen-3-ol, <i>p</i> - cimene, <i>trans</i> - β - ocimene, γ -terpinene, α -terpinolene, <i>cis</i> - <i>p</i> -	NI NI	0.12 2.00	NI NI

				menth-2-en-1-ol, <i>trans-p</i> -menth-2-en-1-ol, borneol, terpin-4-ol, α -terpineol, trans-piperitol, thymol methyl ether, carvacrol methyl ether, thymol, geranyl acetate, <i>trans</i> -caryophyllene, germacrene D, β -bisabolene, espatulenol, caryophyllene oxide, α -eudesmol.			
92	<i>Origanum applii</i> (Lamiaceae) [Sartoratto <i>et al.</i> , 2004]	Herb	AC	Essential Oil: Linalool, terpin-4-ol, cimen-8-ol, thymol methyl ether, carvacrol methyl ether, thymol, β -bourbonene, β -gurjunene, <i>allo</i> -aromadendrene, germacrene D, β -bisabolene, γ -cadinene, espatulenol, α -muurolol, α -cadinol.	NI	>2.0	NI
93	<i>Aloysia tryphila</i> (Verbenaceae) [Sartoratto <i>et al.</i> , 2004]	Herb	AC	Essential Oil: 1-octen-3-ol, p-cimene, limonene, trans- β -ocimene, γ -terpinene, linalool, α -terpineol, neral, geranial, geranyl acetate, cedrene, <i>trans</i> -caryophyllene, curcumene, zingiberene, δ -cadinene, espatulenol, caryophyllene oxide.	NI	0.80	NI
94	<i>Pelargonium graveolens</i> (Geraniaceae) [Hammer <i>et al.</i> , 1999]	Herb	AC	Essential Oil	NI	0.12	NI
95	<i>Pimpinella anisum</i> (Apiaceae)	Seed	AC	Essential Oil	NI	0.5	NI

	[Hammer <i>et al.</i> , 1999]						
96	<i>Pimenta racemosa</i> (Myrtaceae) [Hammer <i>et al.</i> , 1999]	Leaf	AC	Essential Oil	NI	0.12	NI
97	<i>Pinus sylvestris</i> (Pinaceae) [Hammer <i>et al.</i> , 1999]	Needles	AC	Essential Oil	NI	2	NI
98	<i>Piper nigrum</i> (Piperaceae) [Hammer <i>et al.</i> , 1999]	Berry	AC	Essential Oil	NI	>2	NI
99	<i>Pogostemon patchouli</i> (Lamiaceae) [Hammer <i>et al.</i> , 1999]	Leaf	AC	Essential Oil	NI	0.5	NI
100	<i>Prunus armeniaca</i> (Rosaceae) [Hammer <i>et al.</i> , 1999]	Seed	AC	Essential Oil	NI	>2	NI
101	<i>Prunus dulcis</i> (Rosaceae) [Hammer <i>et al.</i> , 1999]	Seed	AC	Essential Oil	NI	>2	NI
102	<i>Thymus vulgaris</i> (Lamiaceae) [Hammer <i>et al.</i> , 1999; Sartoratto <i>et al.</i> , 2004]	Herb	AC	Essential Oil: 1-octen-3-ol, <i>p</i> - cymene, γ -terpinene, linalool, borneol, terpin-4-ol, thymol methyl ether, thymol, carvacrol, <i>trans</i> - caryophyllene, γ - cadienene	NI NI	0.12 2.00	NI NI
103	<i>Salvia sclarea</i> (Lamiaceae) [Hammer <i>et al.</i> , 1999]	Herb	AC	Essential Oil	NI	>2	NI
104	<i>Santalum album</i> (Santalaceae) [Hammer <i>et al.</i> , 1999]	Wood	AC	Essential Oil	NI	0.06	NI
105	<i>Vetiveria zizanioides</i> (Poaceae) [Hammer <i>et al.</i> , 1999]	Leaf	AC	Essential Oil	NI	0.12	NI
106	<i>Zingiber officinale</i> (Zingiberaceae) [Hammer <i>et al.</i> , 1999]	Rhizome	AC	Essential Oil	NI	>2	NI
107	<i>Echinophora platyloba</i> (Umbelliferae) [Avijgan <i>et al.</i> , 2006; Nourozi <i>et al.</i> , 1989*]	Aerial part	Ethanol	(Flavonoid, alkaloid & saponin)*	2	NI	NI
108	<i>Phlomis pungens</i> var. <i>pungens</i> (Lamiaceae)	Dried plant	Chloroform	NI	9	NI	NI

	[Dogan <i>et al.</i> , 2010]						
109	<i>Phlomis armeniaca</i> (Lamiaceae) [Dogan <i>et al.</i> , 2010]	Dried plant	Chloroform	NI	15	NI	NI
110	<i>Tanacetum argenteum</i> (Asteraceae) [Dogan <i>et al.</i> , 2010]	Dried plant	Chloroform	NI	12	NI	NI
111	<i>Pogostemon parviflorus</i> (Lamiaceae) [Najafi <i>et al.</i> , 2011]	Leaf	Methanol Ethanol	Saponins, reducing sugars, tannins, phenols, proteins, triterpenes	10-18 8-15	2.5-10 2.5-20	NI NI
112	<i>Albizia lebbeck</i> (Fabaceae) [Maji <i>et al.</i> , 2010; El- Hawary <i>et al.</i> , 2011*]	Leaf	Water Acetone Benzene	(β -Carotene, linoleic acid, β -sitosterol, amino acids, tannins)*	7 13 6	0.60 0.41 0.50	0.70 0.55 0.65
113	<i>Cleistanthus collinus</i> (Phyllanthaceae) [Maji <i>et al.</i> , 2010; Govindachari <i>et al.</i> , 1969*]	Leaf	Water Acetone Benzene	(Ellagic acid, diphyllin, two lignin lactones, cleistanthin and collinusin)*	8 13 10	0.60 0.39 0.53	0.75 0.450.70
114	<i>Embllica officinalis</i> (Phyllanthaceae) [Maji <i>et al.</i> , 2010; Bhattacharya <i>et al.</i> , 1999*]	Leaf	Water Acetone Benzene	(Embllicanin A,B, punigluconin, pedunculagin)*	9 12 10	0.59 0.48 0.53	0.65 0.55 0.75
115	<i>Eucalyptus deglupta</i> (Myrtaceae) [Maji <i>et al.</i> , 2010]	Leaf	Water Acetone Benzene	NI	4 10 9	- 0.47 0.60	- 0.55 0.85
116	<i>Eupatorium odoratum</i> (Asteraceae) [Maji <i>et al.</i> , 2010]	Leaf	Water Acetone Benzene	NI	8 10 13	0.60 0.50 0.45	- 0.55 0.85
117	<i>Oxalis corniculata</i> (Oxalidaceae) [Maji <i>et al.</i> , 2010]	Leaf	Water Acetone Benzene	NI	6 4 7	- - 0.60	- - 0.65
118	<i>Hevea brasiliensis</i> (Euphorbiaceae) [Maji <i>et al.</i> , 2010]	Leaf	Water Acetone Benzene	NI	8 12 9	0.58 0.58 0.60	0.80 0.70 0.80
119	<i>Lantana camara</i> (Verbenaceae) [Maji <i>et al.</i> , 2010; Ganjewala <i>et al.</i> , 2009*]	Leaf	Water Acetone Benzene	(Carbohydrates, glycolipids, phospholipids, alkaloids, phenolics, terpenoids, phycobatanins, steroid, saponin, phytosterol, tannin)*	10 13 6	0.59 0.53 0.60	0.90 0.60 0.90
120	<i>Adina cordifolia</i>	Leaf	Water	NI	5	-	-

	(Rubiaceae) [Maji <i>et al.</i> , 2010]		Acetone Benzene		7 6	0.80 -	0.90 -
121	<i>Aegle marmelos</i> (Rutaceae) [Maji <i>et al.</i> , 2010]	Leaf	Water Acetone Benzene	NI	4 4 4	- - -	- - -
122	<i>Asparagus racemose</i> (Asparagaceae) [Maji <i>et al.</i> , 2010]	Leaf	Water Acetone Benzene	NI	4 6 5	- - -	- - -
123	<i>Butea frondosais</i> (Fabaceae) [Maji <i>et al.</i> , 2010]	Leaf	Water Acetone Benzene	NI	4 7 4	- 0.78 -	- 1 -
124	<i>Curculigo orchoides</i> (Hypoxidaceae) [Maji <i>et al.</i> , 2010]	Leaf	Water Acetone Benzene	NI	4 4 4	- - -	- - -
125	<i>Dillenia pentagyna</i> (Dilleniaceae) [Maji <i>et al.</i> , 2010]	Leaf	Water Acetone Benzene	NI	5 5.5 4	- - -	- - -
126	<i>Lygodium pinnatifidum</i> (Lygodiaceae) [Maji <i>et al.</i> , 2010]	Leaf	Water Acetone Benzene	NI	4 7 4	- 0.80 -	- 1 -
127	<i>Melastoma malabathricum</i> (Melastomataceae) [Maji <i>et al.</i> , 2010]	Leaf	Water Acetone Benzene	NI	4 8 7	- 0.79 -	- 0.90 -
128	<i>Ocimum basilicum</i> (Lamiaceae) [Maji <i>et al.</i> , 2010]	Leaf	Water Acetone Benzene	NI	4 12 4	- 0.60 -	- 0.85 -
129	<i>Terminalia arjuna</i> (Combretaceae) [Maji <i>et al.</i> , 2010]	Leaf	Water Acetone Benzene	NI	5 7 5	- 0.76 -	- 1 -
130	<i>Valeriana wallichii</i> (Valerianaceae) [Maji <i>et al.</i> , 2010]	Leaf	Water Acetone Benzene	NI	4 4 4	- - -	- - -
131	<i>Thymus mongolicus</i> (Lamiaceae) [Omran <i>et al.</i> , 2009]	Leaf	Hexane	NI	NI	<6.4	NI
132	<i>Mentha pulegium</i> (Lamiaceae) [Omran <i>et al.</i> , 2009]	Leaf	Hexane	NI	NI	<6.4	NI
133	<i>Citrus lemon</i> (Rutaceae) [Omran <i>et al.</i> , 2009]	Leaf	Hexane	NI	NI	<6.4	NI
134	<i>Boesenbergia pandurata</i> (Zingiberaceae) [Taweechaissupapong <i>et al.</i> , 2010]	Yellow rhizome	Ethanol (Maceration) Hydro-distillation	NI Essential Oil: Geraniol (olefinic terpene), linalool, α -	- 15.84	- 0.5	1.25 0.5

				terpineol			
135	<i>Piper sarmentosum</i> (Piperaceae) [Taweechaissupapong <i>et al.</i> , 2010; Masuda <i>et al.</i> , 1991*]	Leaf	Ethanol	(1-allyl-2,6-dimethoxy-3,4-methylenedioxybenzene, 1-allyl-2,4,5-trimethoxybenzene, 1-(1-E-propenyl)-2,4,5-trimethoxybenzene and 1-allyl-2-methoxy-4,5-methylenedioxybenzene)*	10.81	-	1.25
			Hydro-Distillation	Oils	-	-	-
136	<i>Murraya koenigii</i> (Rutaceae) [Zachariah <i>et al.</i> , 2009]	Stem bark	Alkaloids Coumarin	NI	12-19 21	NI NI	NI NI
			Chloroform		11	NI	NI
			Alcohol Acetone		18 20	NI NI	NI NI
137	<i>Embelia ribes</i> (Myrsinaceae) [Suthar <i>et al.</i> , 2009]	Fruits	Petroleum ether Diethyl ether Methanol Water	Benzoquinone derivative: Embelin (2,5-dihydroxy-3-undecyl-2, 5-cyclohexadiene, 4-benzoquinone) and Vilangin	NI NI NI NI	290 660 120 2140	NI NI NI NI
138	Traditional Chinese Medicine (Pseudolaric acid B, Gentiopicrin. Rhein and Alion) [Yan <i>et al.</i> , 2012]	AC	DMSO	Pseudolaric acid B, Gentiopicrin. Rhein and Alion	-	42.22	42.22
139	<i>Morinda citrifolia</i> (Rubiaceae) [Jainkittivong <i>et al.</i> , 2009; Deng <i>et al.</i> , 2010*; West <i>et al.</i> , 2011*]	Fruit	Aqueous	Iridoids (Deacetylasperulosidic acid and Asperulosidic acid)*	NI	40	NI

Abbreviations: Activity not found (-); Not investigated (NI); Available Commercially (AC); Zone of Inhibition in millimetre (ZOI mm); Minimum Inhibitory Concentration for 50 % inhibition (MIC₅₀); Minimum Fungicidal concentration for 50 % sensitivity (MFC₅₀);. (*) in Phytoconstituent column to the respective (*) for reference in same row.

CONCLUSION

Candida albicans is the most frequent pathogenic species followed by *C. glabrata*. *Candida* species associated morbidity and mortality is reported to increase at an alarming rate in India and overseas due to its lead role in hospital acquired infections. Increasing reports on *Candida* resistance to existing antibiotics & invasive Candidosis have led the attention of many researchers to develop alternative plant based effective therapeutics.

A total of 139 plants are listed here for their properties against *Candida albicans* particularly. Appreciable anti-*Candida* properties are observed in members of the families Apiaceae, Lamiaceae, Asteraceae, Fabaceae, Bignomiaceae, Piperaceae, Zingiberaceae, Rutaceae, Poaceae, Cucurbitaceae, Meliaceae, Lauraceae, Verbenaceae, Rutaceae, Rubiaceae. Extracts from *Acacia sp.* (Fabaceae), *Aloe vera* (Xanthorrhoeaceae), *Cinnamomum zeylanicum* (Lauraceae), *Warburgia salutaris* (Canellaceae), *Syzygium aromaticum* (Myrtaceae), *Piper capense* (Piperaceae) and *Cassia tora* (Caesalpinaceae) are reported to exhibit best

characteristic features of inhibiting *Candida albicans* in agar and broth media.

Studies reporting above plants (listed in the table above) as a source of effective anti Candidal drug lacks detailed experimentation that is required for thorough characterization of herbal extracts as per regulatory guidelines. Active phytoconstituents are needed to be isolated and screened for its anti-*Candida* properties like inhibition of yeast to hyphae transition, adhesion and biofilm formation mainly.

Different extracts and essential oils from the various plants are very effective to inhibit the growth of *C. albicans* in the initial experiments but they needs to be explored in adequate details in animal models along with the toxicity studies for establishing them as therapeutic modules against *Candida* species. Because of plant origin, such herbal drugs would be better choices against Candidosis for being safe, effective and inexpensive.

REFERENCES

- Ababutain, I.M. (2011). Antimicrobial Activity of Ethanolic Extracts From some medicinal

- Plants. Australian J of Basic and Applied Sciences. 5(11): 678-683.
- Adhikary, R. and Joshi, S. (2011). Species distribution and anti fungal susceptibility of Candidaemia at a multi super-speciality in southern India. Indian J of Medical Microbiology. 29 (3): 309-11.
- Alimirante, B., Rodriguez, D., and Park, B.J. (2005). Epidemiology and predictors of mortality in cases of *candida* bloodstream infection: Results from population based surveillance, Barcelona, Spain from 2002 to 2003. J. Clin. Microbiol. 43: 1829-1835.
- Anand, J., Rai, N., Kumar, N. and Gautam, P. (2012). Green Tea: A magical herb with miraculous outcomes. International Research Journal of Pharmacy. 3 (5): 139-148.
- Ankri, S. and Mirelman, D. (1999). Antimicrobial properties of Allicin from garlic. Microbes Infect. 1: 125-9.
- Arendrup, M.C., Fursted, K. and Gahrn-Hansen, B. (2005). Semi National surveillance of fungemia in Denmark: Notably high rates of fungemia and numbers of isolates with reduced azole susceptibility. J. Clin. Microbiol. 43: 4434-4440.
- Arya, V., Yadav, S., Kumar, S and Yadav, J.P. (2010). Antimicrobial Activity of *Cassia occidentalis*L. (Leaf) against various Human Pathogenic Microbes. Life Sciences and Medicine Research, 2010: 1-11.
- Avijgan, M., Hafizi, M., Saadat. M., and Nilforoushzadeh, M.A. (2006). Antifungal Effect of *Echinophora platyloba's* Extract against *Candida albicans*. Iranian Journal of Pharmaceutical Research. 4: 285-289.
- Baddley, J.W., Smith, A.M., Moser, S.A. and Pappas, P.G. (2001). Trends in frequency and susceptibilities of *Candida glabrata* bloodstream isolates at a university hospital. Diagnostic Microbiol. Infect. Dis.39: 199-201.
- Bakkali, F., Averbeck, S., Averbeck, D., Zhiri, A., Baudoux, D., and Idaomar, M. (2005). Cytotoxicity and gene induction by some

- essential oils in the yeast *Saccharomyces cerevisiae*. Mutation Research. 585: 1-3.
- Beckman, J. (2007). Antimicrobial properties of Pomegranate fruit: A thesis. Willamette University.
- Bhattacharya, A., Chatterjee, A., Ghosal, S., and Bhattacharya, S.K. (1999). Antioxidant activity of active tannoid principles of *Embllica officinalis* (amla). Indian I. Exp. Biol. 37: 676-680.
- Bhattarai, N., and Karki, M. (2004). Medicinal and aromatic plants - Ethnobotany and conservation status. In: J. Burley, J. Evans and J. Youngquist (Eds.). Encyclopedia of Forest Sciences. Academic Press, London, UK. 523-532.
- Chakravarthi, S., Wei, C.Z., Nagaraja, H.S., Fung, W.S., Wah, M.J., and Sreekumar, S. (2010). A histopathological study of severity of cerebral candidiasis as a result of immunosuppression caused by breast carcinoma. Int. J. Mol. Med. Adv. Sci. 6: 1-7.
- Colombo, A.L., Melo, A.S.A., Rosasa, R.F.C., Salmao, R., and Briones, M. (2003). Outbreak of *Candida rugosa* Candidemia: An emerging pathogen that may be refractory to amphotericin B therapy. Diagn. Microbiol. Infect. Dis.46: 253-257.
- Colombo, A.L., Nucci, M., and Park, B.J. (2006). Epidemiology of candidemia in Brazil: A nationwide sentinel surveillance of candidemia in eleven medical centres. J. Clin. Microbiol. 44: 2816-2823.
- Dalirsani, Z., Adibpour, M., Aghazadeh, M., Amirchaghmaghi, M., Falaki, F., Mozafari, P.M., and Hamzei, F.M. (2011). In vitro Comparison of Inhibitory Activity of 10 Plant Extracts Against *Candida albicans*. Australian J of Basic and Applied Sciences. 5(5): 930-935.
- Damke, E., Tsuzki, J.K., Cortez, D.A.G., Ferreira, I.C.P., Bertoni, T.A., Batista, M.R., Donati, L., Svidzinski, T.E.I., and Consolaro, M.E.L. (2011). *In vivo* activity of *Sapindussaponaria* against azole-susceptible and –resistant

- human vaginal *Candida* species. BMC Complementary and Alternative Medicine. 11:35.
- Deng, S., West, B.J., Palu, A.K. and Jensen, C.J. (2010). Determination and comparative analysis of major iridoids in different parts and cultivation sources of *Morinda citrifolia*. *Phytochem., Analysis*, 22(1): 26-30.
- Dogan, N.M., Cansaran, A., Acar, G., and Oztekin, M. (2010). Antimicrobial Activity of Extracts of some Plants from Amasya (Turkey). *Advances in Bio research*. 1(1): 87-91.
- El-Hawary, S., El-Fouly, K., Sokkar, N.M., and Talaat, Z. (2011). A phytochemical profile of *Albizia lebbbeck* (L.) Benth. Cultivated in Egypt. *Asian J. Of Biochem*. 6 (2): 122-141.
- Endo, E.H., Cortez, D.A.G., Nakamura, T.U., Nakamura, C.V., and Filho, B.P.D. (2010). Potent antifungal activity of extracts and pure compounds isolated from pomegranate peels and synergism with fluconazole against *Candida albicans*. *Science Direct: Research in Microbiology*. 161: 534-540.
- Fidel, Jr. P.L. (2006). *Candida* host interactions in HIV disease: Relationships in oropharyngeal candidiasis. *Adv. Dental res*. 1: 80-84.
- Ganjewala, D., Sam, S., and Khan, K.H. (2009). Biochemical composition and antibacterial activities of *Lantana camara* plants with yellow, lavender, red and white flowers. *Eur. Asia J. Biosci*. 3: 69-77.
- Govindachari, T.R., Sathe, S.S., Viswanathan, N., Pai, B.R., and Srinivasan, M. (1969). Chemical constituents of *Cleistanthus collinus* (Roxb.). *Tetrahedron*. 25: 2815-2821.
- Gudlaugsson, O., Gillespie, S., and Lee, K. (2003). Attributable mortality of nosocomial candidemia, revisited. *Clin. Infec. Dis*. 37: 1172-1177.
- Hajjeh, R.A., Safair, A.N., and Harrison, L.H. (2004). Incidence of bloodstream infections due to *Candida* species and *in vitro* susceptibilities of isolates collected from

- 1998 to 2000 in a population-based active surveillance program. *J. Clin. Microbiol.* 42: 1519-1527.
- Hamman, J.H. (2008). Composition and Application of Aloe vera leaf gel. *Molecule.* 13: 1599-1616.
- Hammer, K.A., Carson, C.F., and Riley, T.V. (1999). Antimicrobial activity of essential oils and other Plant Extracts. *Journal of Applied Microbiology.* 86: 985-990.
- Hoffling, J.F, Anibal, P.C., Obando-Pereda, G.A., Peixoto, I.A.T., Furletti, V.F., Foglio, M.A., and Gonçalves, R.B. (2010). Antimicrobial potential of some plant extracts against *Candida* species. *Braz. J. Biol.* 70(4): 1065-1068.
- Ibrahim, M., Srinivas, M., and Lakshmi, N.M. (2011). Phyto-chemical analysis and Anti-microbial evaluation of Aloe vera gel against some Human and Plant pathogens. *Asian J of Current Chemistry.* 1(1): 1-11.
- Jainkittivong, A., Butsarakamruha, T., and Langlais, R.P. (2009). Antifungal Activity of *Morinda citrifolia* fruit extract against *C. albicans*. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology Endodontology.* 108(3): 394-398.
- Jebashree, H.S., Kingsley, S.J., Sathish, E.S., and Devapriya, D. (2011). Antimicrobial Activity of Few Medicinal Plants against Clinically Isolated Human Carciogenic Pathogens: An *In vitro* Study. *ISRN Dentistry.* 2011: 1-6.
- Joshi, S., Gupta, P., Kumar, N., Rai, N., Gautam, P., and Thapliyal, A. (2013). A comprehensive report on therapeutic potential of *Elaeocarpus ganitrus* Roxb. (Rudraksha). *Environment Conservation Journal.* 13(3): 147-150.
- Kothari A. and Sagar A. (2009). A report on Epidemiology on *Candida* bloodstream infections in a tertiary care institute in India. *Indian Journal of Medical Microbiology* 27(2): 17-178.
- Khan, R., Islam, B., Akram, M., Shakil, S., Ahmad, A., Ali, S.M., Siddiqui, M., and Khan, A.U. (2009). Antimicrobial Activity of Five

- Herbal Extracts against Multi Drug Resistant Strains of Bacteria and Fungus of Clinical Origin. *Open Access Molecules*. 14: 586-597.
- Kumar, A., Bhatti, V., Kumar, A., Patil, S., Bhatia, V., and Kumar, A. (2011). Screening of Various Plant Extracts for Antifungal Activity against *Candida* Species. *World J of Science and Technology*. 1(10): 43-47.
- Kumar, A., Rai, N., Kumar, N., Gautam, P., and Kumar, J.S. (2013). Mechanisms involved in hepatoprotection of different herbal products: A Review. *Int. J. Res. Pharm. Sci*. 4(2), 1-6.
- Lewington, A. (2003). Medicinal plants and plant extracts: A review of their importation into Europe. *Traffic International*, Cambridge, UK.
- Maji, S., Dandapat, P., Ojha, D., Maity, C., Halder, S.K., and Mohapatra, P.K.D., *et al*. *In vitro* Antimicrobial Potentialities of Different Solvent Extracts of Ethnomedicinal Plants against Clinically Isolated Human Pathogens. *Journal of Phytology*, 2010; 2(4): 57-64.
- Malani, P.N., Bradley, S.F., Little, R.S., and Kauffman, C.A. (2001). Trends in species causing fungemia in a tertiary care medical centre over 12 years. *Mycoses*. 44: 446-449.
- Marchetti, O., Bille, J., and Flukiger, U. (2004). Epidemiology of candidemia in Swiss tertiary care hospitals: Secular trends, 1991-2000. *Clin. Infect. Dis*. 38: 311-320.
- Marr, K.A., Seidel, K., White, T.C., and Bowde, R.A. (2000). Candidemia in allogeneic blood and marrow transplant recipients: Evolution of risk factors after the adaptation of prophylactic flucanazole. *J. Infect. Dis*. 181: 309-316.
- Masuda, T., Inzumi, A., Yamada, Y., Padolina, W.G., Kikuzaki, H., and Nakatani, N. (1991). Antimicrobial phenylpropanoids from *Piper sarmentosum*. *Phytochemistry*. 30: 3227-3228.
- Michalopoulos, A.S., Geroulanos, S., and Mentzelopoulos, S.D. (2003). Determinants of Candidemia and candidemia-related deaths

- in cardiothoracic ICU patients. *Chest*. 124: 2244-2255.
- Mwambete, K.D. (2009). The *in vitro* antimicrobial activity of fruit and leaf crude extracts of *Momordica charantia*: A Tanzanian Medicinal Plant. *African Health Sciences*. 9: 34-39.
- Nagle, D.G., and Zohu, Y.D. (2006). Natural Product-Derived Small Molecule Activators of Hypoxia-Inducible Factor-1 (HIF-1). *Curr Pharm Des*. 12(21): 2673–2688.
- Najafi, S., and Sadeghi-Nejad, B. (2011). Screening of *Pogostemon parviflorus* Benth. For Anti-*Candida* activity. *African Journal of Microbiology Research*. 5(6): 657-660.
- Nourozi, M. (1989). Evaluation of Phytochemical and Antimicrobial effect of *Echinophora platyloba*. Pharm. D. Thesis. Tehran University of Medical Sciences, Tehran. 35-40.
- Okerele O (1992). WHO Guidelines for the Assessment of Herbal Medicines. *Fitoterapia*. 63 (2): 99-110.
- Omran, S.M., and Esmailzadeh, S. (2009). Comparison of anti-*Candida* activity of Thyme, Pennyroyal, and Lemon Essential oils versus Antifungal Drugs against *Candida* species. *Jundishapur Journal of Microbiology*. 2(2): 53-60.
- Pai, M.B.H., Prashant, G.M., Murlikrishna, K.S., Shivakumar, K.M., and Chandu, G.N. (2010). Antifungal efficacy of *Punicagranatum*, *Acacia nilotica*, *Cuminumcyminum* and *Foeniculumvulgareon Candida albicans*: An *in vitro* study. *Indian J. Dent. Res*. 21(3): 223-231.
- Panda, N.P., and Ray, P. (2012). A study on Effect of Some Indigenous Plant extracts against Two Human Pathogens. *Asian J. Exp. Biol. Sci*. 3(1): 175-179.
- Pappas, P.G., Rex, J.H., and Lee, J. (2003). A Prospective observational study of candidemia: Epidemiology, therapy and influences on mortality in hospitalised adult

- and paediatric patients. Clin. Infect. Dis. 37: 634-643.
- Prabhakar, K., Kumar, L.S., Rajendran, S., Chandrasekaran, M., Bhaskar, K., and Khan, K.S. (2008). Antifungal Activity of Plant Extracts against *Candida* Species from Oral Lesions. Indian Journal of Pharmaceutical Sciences. 2008: 801-803.
- Rai, N., Kumar, N., Gautam, P., and Rawat, P. (2012). Herbal plants as potent candidate for Anti-ulcer drug development. Environment Conservation Journal. (1 & 2): 187-189.
- Rangel-Frausto, M.S., Wiblin, T., and Blumberg, H.M. (1999). National epidemiology of mycoses survey (NEMIS): Variations in rate of bloodstream infections due to *Candida* species in seven surgical intensive care units and six neonatal care units. Infec. Dis. 29: 253-25.
- Redding, S., Smith, J., Farinacci, G., Rinaldi, M., Fothergill, A., Rhine-Chalberg, J., and Pfaller, M. (1993). Resistance of *Candida albicans* to Fluconazole during treatment of Oropharyngeal Candidiasis in a Patient with AIDS: Documentation by in vitro Susceptibility testing and DNA Subtype Analysis. Clinical Infect. Dis. 18(2): 240-242.
- Richet, H., Roux, P., Des Champs, C., Esnault, Y., and Andremont, A. (2002). Candidemia in French hospitals; Incidence rates and characteristics. Clin. Microbiol. Infection. 8: 405-412.
- Ruwali, P., Rai, N., Kumar, N., and Gautam, P. (2013). Antiviral potential of medicinal plants: An Overview. International Research Journal of Pharmacy. 4 (6), 8-16.
- Saad, B., Azaizeh, H., and Said, O. (2005). Tradition and Perspectives of Arab herbal medicine: A review. Evidence-Based Complementary and Alternative Medicine. 2(4): 475-479.
- Samei, A., Tambani, T., Harshfield, E., Ramalivhana, J.N., and Bessong, P.O. (2010). Antifungal activities of Selected Venda medicinal plants against *Candida albicans*,

- Candida krusei* and *Cryptococcus neoformans* isolated from South African AIDS Patients. *African J of Biotechnology*. 9: 2965-2976.
- Sandven, P., Bevanger, L., Digrahes, A., Haukland, H.H., Mannsaker, T., and Gaustad, P. (2008). Candidemia in Norway (1991 to 2003): Results from a nationwide study. *J. Clin. Microbiol.* 44: 1977-1981.
- Sanglard, D., and Odds, F.C. (2002) Resistance of *Candida* species to antifungal agents: Molecular mechanisms and clinical consequences. *Lancet Infect. Dis.* 2:73-85.
- Sartoratto, A., Machado, A.L.M., Delarmelina, C., Figuera, G.M., Duarte, M.C.T., and Rehder, V.L.G. (2004). Composition and Antimicrobial Activity of Essential oils from Aromatic Plants used in Brazil. *Brazilian Journal of Microbiology*. 34: 275-280.
- Singh, H.B., Srivastava, M., Singh, A.B., and Srivastava, A.K. (1995). Cinnamon bark oil, a potent fungitoxicant against fungi causing respiratory tract mycoses. *Allergy*. 50: 995-9.
- Suthar, M., Patel, R., Hapani, K., and Patel, A. (2009). Screening of *Embelia ribes* for Antifungal activity. *International J of Pharmaceutical Sciences and drug Research*. 1(3): 203-206.
- Suvarna, V., and Patil, S. (2009). Antifungal activity of Selected Plants Extracts against Human Fungal Pathogens. *Journal of Herbal Medicine and Toxicology*. 3(2): 151-153.
- Takakura, S., Fujhhara, N., Saito, T., Kudo, T., Iinuma, Y., and Ichiyana, S. (2004). National surveillance of species distribution in blood isolates of *Candida* species in Japan and their susceptibility to six antifungal agents including voriconazole and micafungin. *J. Antimicrobial Chemother.* 53: 283-289.
- Taweechaissupapong, S., Singhara, S., Lertsatitthanakorn, P., and Khunkitti, W. (2010). Antimicrobial effects of *Boesenbergia pandurata* and *Piper sarmentosum* leaf extracts on planktonic cells and biofilm of oral pathogens. *Pak J Pharm Sci.* 23(2): 224-231.

- Tzuzuki, J.K., Svidzinski, T.I.E., Shinobu, C.S., Silva, L.F.A., Rodrigues-Filho, E., Cortez, D.A.G., and Ferreira, I.C.P. (2007). Antifungal activity of the extracts and saponins from *Sapindus saponaria* L. *Anais Ac Bras Ciências*.79:577-583.
- Voss, A., Kluytmans, J.A., and Koelman, J.G. (1996). Occurrence of yeast bloodstream infections between 1987 to 1995 in five Dutch University hospitals. *Eur. J. Clin. Microbiol. Infect. Dis*.15: 909-912.
- Wang, H., and Ng, T.B. (2000). Purification of lillin a novel antifungal protein from bulbs of round-clove garlic, onion. *Life sci*. 75: 211-15.
- Wang, H.X. and Ng, T.B. (2001). Purification of allivin a novel antifungal protein from bulbs of the round-clove garlic. *Life sci*. 70: 357-65.
- West, B.J., Deng, S. and Jensen, C.J. (2011). Nutrient and phytochemical analyses of processed noni puree. *Food Res. Int.*, 44(7): 2295-2301.
- Wey, S.B., Mori, M., Pfaller, M.A., Woolsen, R.F., and Wenz, R.P. (1988). Hospital-acquired candidemia; the attributable mortality and excess length of stay. *Arch. Internal Med*.148: 2642-2645.
- WHO. 1978. The Promotion and Development of Traditional Medicine. WHO Technical Report Series, No. 622:8, Geneva, Switzerland.
- Wisplinghoff, H., Bischoff, T., Tallent, S.M, Seifert, H., Wenzel, R.P., and Edmond, M.B. (2004). Nosocomial bloodstream infections in US hospitals: Analysis of 24,179 cases from a prospective nationwide surveillance study. *Clin. Infect. Dis*. 39: 309-317.
- Xia, L., and Ng, T.B. (2005). Isolation of alliumin, a novel protein with antimicrobial and antiproliferative activities from multiple-cloved garlic bulbs. *Peptides*. 26: 177-83.
- Yan, Z., Hua, H., Xu, Y., and Samaranayake, L.P. (2012). Potent anti-fungal Activity of Pure Compounds from Traditional Chinese

Medicine Extracts against Six Oral *Candida* spp. Hindawi Publishing Corporation. 2011: 1-8.

species and the synergy with Fluconazole against azole resistant *Candida albicans*. Hindawi Publishing Corporation, Evidence Based Complementary and Alternative Medicine. 2012: 1-6.

Yebpella, G.G., Hammuel, C., Adeyemi, H.M.M., Magomya, A.M., Agbaji, A.S. and Shallangwa, G.A. (2011). Phytochemical Screening and comparative study of antimicrobial activity of Aloe vera various extracts. African J of Microbiology Research, 5(10): 1182-1187.

Zachariah, S.M., Muthumani, and Ramaseshu, K.V. (2009). Phytochemistry and Antimicrobial Screening of Stem Bark of *Murraya koenigii*(Linn) Spreng. The Internat. Journal of Pharmacology. 2009: 6.

Zeng, H., Tian, J., Zheng, Y., Ban, X., Zeng, J., Mao, Yehong. (2011). In vitro and In vivo Activities of Essential Oil from Seeds of *Anethems graveolens* L. against *Candida*